

Combining benzo[*d*]isosenazol-3-ones with sterically hindered alicyclic amines and nitroxides: enhanced activity as glutathione peroxidase mimics

Tamas Kálai,^a Govindasamy Mugesh,^b Gouriprasanna Roy,^b Helmut Sies,^c Zoltán Berente^d and Kálmán Hideg^{*a}

^a Institute of Organic and Medicinal Chemistry, University of Pécs, P. O. Box 99, H-7602, Pécs, Hungary. E-mail: kalman.hideg@aok.pte.hu; Fax: +36-72-536219; Tel: +36-72-536221

^b Department of Inorganic and Physical Chemistry, Indian Institute of Science, Bangalore, 560012, India

^c Institut für Biochemie und Molekularbiologie I, Heinrich-Heine-Universität Düsseldorf, Postfach 101007, D-40001, Düsseldorf, Germany

^d Institute of Biochemistry and Medical Chemistry, University of Pécs, P. O. Box 99, H-7602, Pécs, Hungary

Received 13th July 2005, Accepted 8th August 2005
First published as an Advance Article on the web 31st August 2005

Benzo[*d*]isosenazol-3-ones *N*-substituted with sterically hindered diamagnetic and paramagnetic five- or six-membered nitroxides or their precursors, including ring-opened diselenides, exhibit synergism in glutathione peroxidase (GPx) activity.

Introduction

Glutathione peroxidases (GPx) are selenoenzymes that protect various organisms from oxidative stress by catalyzing the reduction of hydroperoxides at the expense of glutathione (GSH) (eqn 1).^{1,2}



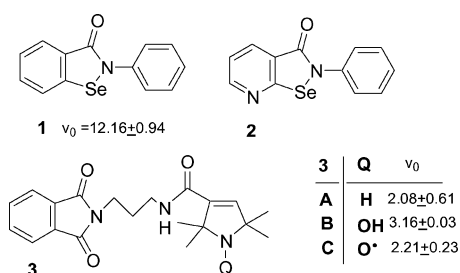
The GPx superfamily contains four types of enzymes, the classical cytosolic GPx (cGPx), phospholipid hydroperoxide GPx (PHGPx), plasma GPx (pGPx) and gastrointestinal GPx (giGPx), all of which require selenium in their active sites for catalytic activity.^{3–9} The biological role of these key enzymes in the antioxidant defense system comprises not only detoxification, by reducing an overproduction of hydroperoxides, but also the regulation of intracellular signalling pathways¹⁰ and enzyme activities, such as that of 5-lipoxygenase.¹¹ To overcome the intrinsic difficulties associated with the use of an enzyme as a drug, a number of low-molecular-weight organoselenium mimics^{12–14} have been developed for the reduction of hydroperoxides, which include the well-known GPx mimic ebselen (**1**)^{15,16} (Scheme 1.) and related compounds having direct Se–N bond,^{17–22} cyclic compounds without any direct Se–N bond^{23,24} α -phenylselenoketones,²⁵ and diaryl diselenides having Se...N or Se...O intramolecular interactions.^{26–30} Ebselen (**1**) is a nontoxic compound at pharmacologically active concentrations, because

its selenium is not bioavailable. It is mostly bound to proteins in the form of selenyl sulfides^{15,16,31} and it is metabolized predominantly into glucuronidated species.³² Another important feature of ebselen is its inability to oxidize GSH in the presence of oxygen which normally leads to the uncontrolled production of superoxide and other free radical species.³³ Although a number of attempts have been made to design and synthesize ebselen-related GPx mimics based on substituent effects or isosteric replacements, most of them met with limited success. For example, the replacement of the phenyl ring in ebselen by a pyridine ring (**2**) resulted in a complete loss of catalytic activity.²⁰ In view of the potential applications of ebselen and related derivatives, we synthesized ebselen-based compounds by incorporating five- or six-membered *N*-heterocycles (as secondary amine connected to a diselenide “A form” or oxidized forms: *N*-hydroxyls “B form”, *N*-oxyls “C form”). A number of non-selenium compounds having these substituents have been previously shown to possess cardioprotective activity.³⁴ It has also been shown that pyrroline-based compounds such as **3A** exhibit markedly enhanced protection against ischemia/reperfusion-induced myocardial contractile dysfunction³⁵ and postischemic myocardial injury³⁶ probably due to their combined antioxidative and antiarrhythmic activities because the *in vivo* oxidation of **3A** to **3C**.³⁷

In this work, we report the GPx activity of a series of closely related benzoselenazolones and show, for the first time, that the synergistic effect of selenium and pyrroline substituents enhances the antioxidant activity of these compounds.

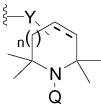
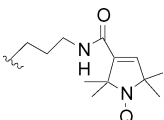
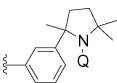
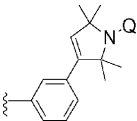
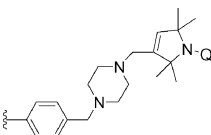
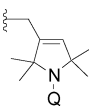
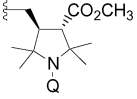
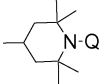
Results and discussion

Compounds **12C**–**18C** were synthesized using the method described earlier,³⁸ *i.e.*, by treating paramagnetic amines **5**–**11** in CHCl₃ with 2-chloroselenyl-benzoylchloride **4**³⁹ at ambient temperature in the presence of 2 eq. Et₃N. Amines **5**,³⁴ **6**,⁴⁰ **9**,⁴¹ **10**⁴² and **11**⁴³ were prepared according to published procedures. Amine **7** was prepared by Suzuki coupling of paramagnetic vinylbromide **19**⁴⁴ with 3-nitrobenzene boronic acid in the presence of Ba(OH)₂ and PdCl₂(PPh₃)₂ as a catalyst in aq. dioxane followed by reduction of the resulted aromatic nitro compound **20** by Ehrenkauer's method.⁴⁵ Alkylation of paramagnetic



Scheme 1 Structure of ebselen **1**, pyridine analogue of ebselen **2**, cardioprotective compounds **3A**–**C** and observed initial reaction rates (v_0 , $\mu\text{M min}^{-1}$) in the reaction of **1**.

Table 1 Initial rates (v_0) for the reduction of *t*-BuOOH in the presence of catalysts^a

| Compound |  | Form (Q), $v_0/\mu\text{M min}^{-1}$ [A: Q=H, B: Q=OH, C: Q=O [•]] |
|----------|---|--|
| 5, 12 |  | 12A 19.18 ± 1.65 12B 5.10 ± 0.95 12C 30.94 ± 0.86 |
| 6, 13 |  | 13A 25.11 ± 0.16 |
| 7, 14 |  | 14A 11.94 ± 0.41 |
| 8, 15 |  | 15A 30.13 ± 0.15 |
| 9, 16 |  | 16A 13.82 ± 0.46 16C 22.11 ± 0.57 |
| 10, 17 |  | 17A 17.46 ± 0.76 17C 14.05 ± 0.76 17C 14.05 ± 0.76 |
| 11, 18 |  | 11A 2.44 ± 0.19 18B 23.57 ± 0.76 18C 21.65 ± 0.38 |

^a Conditions: GSH: 1 mM; DTPA: 1 mM; GSSG reductase: 0.6 unit mL⁻¹; NADPH: 0.1 mM; *t*-BuOOH: 1.2 mM; selenium catalysts: 0.05 mM; in 0.1 M potassium phosphate buffer, pH 7.3, $n = 3$.

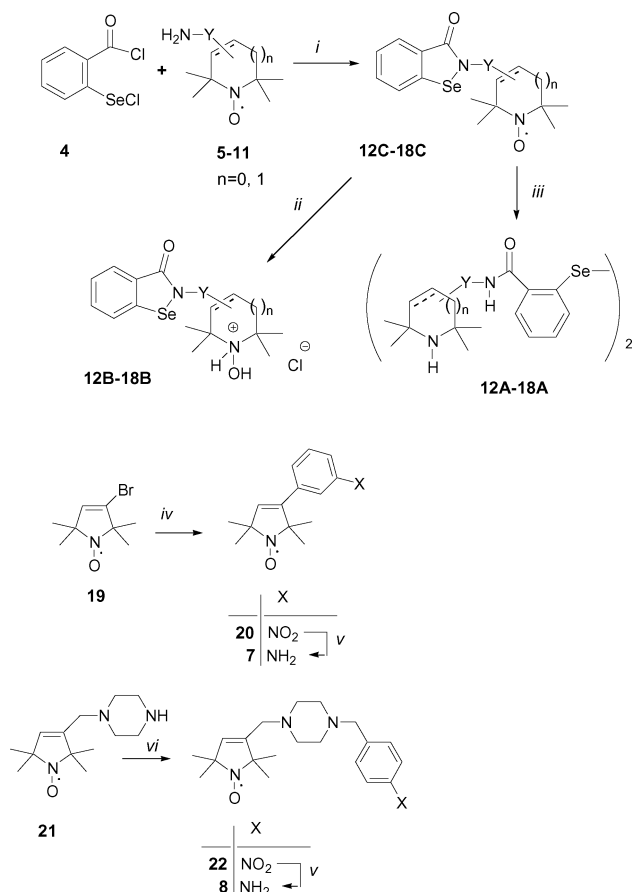
piperazine **21**⁴⁶ with 4-nitrobenzylbromide afforded compound **22** the aromatic nitro group of which was reduced to yield amine **8** with ammonium formate in the presence of palladium on charcoal. The *N*-hydroxylamines **12–18** “B form” was achieved by refluxing the corresponding nitroxides **12–18** “C form” in EtOH saturated with HCl gas.³⁵ Reduction of the “C form”, a nitroxide with Fe powder in glacial acetic acid⁴⁷ yielded the corresponding secondary amine diselenides **12–18** “A form” with opening of isoselenazole ring (Scheme 2).

This ring opening takes place quite easily, even by reduction with ascorbic acid to selenol and oxidation during work-up or standing on air results in formation of a diselenide **23** (Scheme 3).

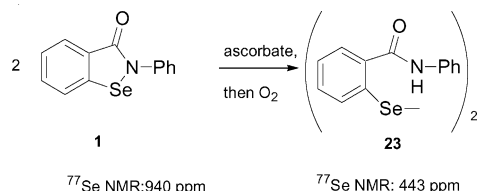
The ring opening followed by oxidation, e.g. structure of **12A–18A** compounds was observed by ⁷⁷Se NMR measurements and data are in good agreement with earlier observations.⁴⁸

The GPx activity of new compounds for reduction of *t*-BuOOH was screened spectrophotometrically at 340 nm as described earlier⁴⁹ with minor modifications. It is evident from data in Scheme 1 and Table 1 that most of the selenium compounds used in the present study are more potent than ebselen (**1**). Interestingly, compounds **3A**, **3B** and **3C** which lack selenium in the benzene anellated five-membered ring and **11** nitroxide precursor, exhibited some GPx activity, however they are about ten-fold less active than isoselenazolone derivatives (Scheme 1). The pyrroline-substituted compounds

showed remarkable GPx activity when attached to the basic benzoselenazolone unit. For example, the activity of compound **12C** ($v_0 = 30.94 \pm 0.86 \mu\text{M} \times \text{min}^{-1}$), in which both pyrroline and isoselenazole units are present, is higher than the sum of their activities in the individual cases [v_0 (**1** + **3C**) = $14.37 \pm 1.17 \mu\text{M} \times \text{min}^{-1}$], suggesting a synergistic effect. The catalytic effect was demonstrated by changing the concentration of **12C** and GSH (Table 1). The observed initial reduction rates (v_0) were directly proportional to the catalyst concentration and rate increases with increasing concentration of GSH and with excess amount of it, the expected saturation kinetics were observed (figure not shown). We can find the same case in the catalytic activity of compound **18C**, which mimics the shape of ebselen with a six-membered ring connected to the nitrogen of isoselenazolone. This compound exhibits higher activity than ebselen (**1**) itself, confirming the role of the nitroxide based substituent. In other words, these two functionalities (isoselenazole and pyrroline/pyrrolidine/piperidine) individually show moderate effects on the reduction rate, however, when they are present together, the effect is supra-additive. The substitution pattern in the pyrroline ring also affects the reduction rate as observed for compounds **12A** and **13A**. Compound **13A**, in which the pyrroline ring is attached to the phenyl ring at the 2-position, exhibits 2-fold higher activity than the 3-substituted **14A**. The best initial rates were observed in the case of compounds with



Scheme 2 . Reagents and conditions: (i) **4** (1.0 eq.), amine **5-11** (1.0 eq.), Et₃N (2.0 eq.), CHCl₃, rt, 1 h, 35–73%; (ii) EtOH–HCl, reflux, 20 min., 68–82%; (iii) Fe, AcOH, 70 °C, 1 h, then K₂CO₃, 47–59%; (iv) 3-nitrophenyl boronic acid (1 eq.), Pd(PPh₃)₂Cl₂ (5%), Ba(OH)₂ (1 eq.), dioxane–water, reflux, 3 h, 44%; (v) HCO₂NH₄ (8 eq.), Pd/C, MeOH, 40 °C, 2 h, 35–43%; (vi) 4-nitrobenzylbromide (1 eq.), K₂CO₃, CHCl₃, reflux, 4 h, 68%.



Scheme 3 Reduction of ebselen with ascorbic acid, followed by a spontaneous oxidation in an NMR tube.

a polar spacer group *e.g.* an amide for **12C** and an amine for **15A**. The combined catalytic effect appears to be less in the case of apolar spacers: **16A** with a methylene group and **14A** with a phenyl group as a spacer have less catalytic activity. It is interesting to note that compounds **18B** and **18C** without linking groups also show similar GPx activity as **16C**. This indicates that the ring size of nitroxide heterocycle does not affect the reaction rate. Seemingly, the oxidation state of nitroxide moiety does not effect the initial reaction rate neither the isoselenozonolone/diselenide form; **12C** is more effective than **12A**, but as efficient as **15A**.

Conclusions

In conclusion, we have shown that due to the synergistic effect of selenium and amino functionalities the modification of the basic ebselen unit by introducing redox-active pyrroline groups greatly enhances the GPx activity of ebselen. The results presented here suggest a new concept that sterically hindered amino groups near the active site of GPx may act synergistically with selenium

during the reduction of hydroperoxides. Compounds like **12-18** with broad antioxidant activity may serve as “ROS and RNS sponges” and can be promising candidates in future therapy of free radical mediated diseases.

Experimental

General

Melting points were determined with a Boetius micro melting point apparatus and are uncorrected. Elemental analyses (C, H, N, S) were performed on Carlo Erba EA 1110 CHNS elemental analyser. The IR (Specord 85) spectra were in each case consistent with the assigned structure. Mass spectra were recorded on a VG TRIO-2 instrument in the EI mode (70 eV, direct inlet), the source temperature was 210 °C, or an with Automass Multi instrument in the EI mode (70 eV, direct inlet). ESR spectra were obtained from 10⁻⁵ molar solutions (CHCl₃), using an MS200 (Magnettech GMBH, Berlin) spectrometer. All radicals exhibited three equally spaced lines with *a_N* = 15.1–15.5 G. ⁷⁷Se NMR spectra were obtained on either a Bruker AVACE400 NMR spectrometer in CDCl₃–MeOH (1 : 1) mixture or on a Varian INOVA 400 WB instrument and chemical shifts are reported with respect to Me₂Se. To obtain high resolution ⁷⁷Se NMR spectra of **12C** it was reduced with the excess of co-dissolved (PhNH)₂. Ebselen and ascorbic acid reaction was recorded in CDCl₃–DMF (1 : 1) mixture. ¹H and ¹³C NMR spectra were recorded with a Varian INOVA 400 WB spectrometer at 400 MHz at 25 °C, chemical shifts are given in ppm. Preparative flash column chromatography was performed on Merck Kieselgel 60 (0.040–0.063 mm). Qualitative TLC was carried out on commercially prepared plates (20 × 20 × 0.02 cm) coated with Merck Kieselgel GF₂₅₄. Compounds **3A–C**,³⁵ **4**,³⁹ **5**,³⁵ **6**,⁴⁰ **9**,⁴¹ **10**,⁴² **13C**,³⁸ **19**,⁴⁴ **22**,⁴⁶ **23**⁴⁸ were prepared according to published procedures, compound **1** was purchased from Calbiochem, compound **11A** and other reagents were purchased from Aldrich. The GPx activity was followed spectrophotometrically at 340 nm on a Perkin-Elmer Lambda 35 UV-VIS Spectrophotometer. The test mixture contained GSH (1 mM), DTPA (1 mM), glutathione disulfide reductase (0.6 unit mL⁻¹), and NADPH (0.1 mM) in 0.1 M potassium phosphate buffer, pH 7.3. GPx samples were added to the test mixture at room temperature and the reaction was started by the addition of *tert*-butyl hydroperoxide (1.2 mM, final concentration). The initial reduction rates were calculated from the rate of NADPH oxidation at 340 nm. Each initial rate was measured at least three times and calculated from the first 5–10% of the reaction by using 6.22 mM⁻¹ cm⁻¹ as the extinction coefficient for NADPH. For the peroxidase activity, the rates were corrected for the background reaction between *t*-BuOOH and GSH.

Synthesis of 3-(3-nitrophenyl)-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-1-yloxy radical (20). A mixture of compound **19** (1.1 g, 5.0 mmol), Ba(OH)₂·8H₂O (1.57 g, 5.0 mmol), 3-nitrophenylboronic acid (830 mg, 5.0 mmol) and PdCl₂(PPh₃)₂ (140 mg, 0.2 mmol) in dioxane–water (24 mL : 6 mL) was stirred and refluxed for 3 h. After cooling the dioxane was evaporated off, the aqueous phase was extracted with CH₂Cl₂ (2 × 20 mL). The organic phase was washed with water (20 mL), separated, dried (MgSO₄), filtered and evaporated under reduced pressure and the residue was purified by flash column chromatography (hexane–EtOAc) on silica gel to give compound **20** as a yellow solid 574 mg (44%), mp 69–71 °C. *R_f*: 0.50 (hexane–EtOAc, 2 : 1). Anal. calc. for C₁₄H₁₇N₂O₃: C 64.35, H 6.56, N 10.72; found: C 64.18, H 6.51, N 10.64%. MS (EI): *m/z* (%) 261 (M⁺, 28), 246 (100), 231 (53), 216 (25), 128 (37).

General procedure for Ehrenkauffer reduction (7, 8)

To a stirred solution of nitro compound **20** or **22** (5.0 mmol) and HCO₂NH₄ (2.52 g, 40 mmol) in MeOH (30 mL) at 40 °C

under N₂ atmosphere 120 mg Pd/C (10%) was added and the mixture was stirred and refluxed until consumption of starting material (ca. 2 h). After cooling, the mixture was filtered through Celite and washed with methanol–water (40 mL : 10 mL). The methanol was evaporated off, the aqueous layer saturated with solid K₂CO₃ and extracted with CHCl₃–MeOH (9 : 1) (2 × 20 mL). The organic phase was dried (MgSO₄), and activated MnO₂ (200 mg) was added and O₂ was bubbled through the solution at rt for 30 min. The mixture was then filtered, evaporated and the residue was purified by flash column chromatography (CHCl₃–MeOH) to give the title compounds.

3-(3-Aminophenyl)-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-1-yloxy radical (7). 404 mg (35%), mp 105–107 °C. *R_f*: 0.30 (CHCl₃–Et₂O, 2 : 1). Anal. calc. for C₁₄H₁₉N₂O: C 72.69, H 8.28, N 12.11; found: C 72.56, H 8.22, N 12.01%. MS (EI): *m/z* (%) 231 (M⁺, 70), 216 (18), 201 (67), 186 (100), 158 (77).

3-[4-(4-Aminobenzyl)piperazin-1-ylmethyl]-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-1-yloxy radical (8). 737 mg (43%), mp 112–115 °C. *R_f*: 0.10 (CHCl₃–MeOH, 9 : 1). Anal. calc. for C₂₀H₃₁N₄O: C 69.92, H 9.10, N 16.32; found: C 69.90, H 9.05, N 16.19%. MS (EI): *m/z* (%) 343 (M⁺, 22), 313 (17), 222 (26), 207 (84), 106 (100).

3-[4-(4-Nitrobenzyl)piperazin-1-ylmethyl]-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-1-yloxy radical (22). A solution of compound **21** (2.38 g, 10.0 mmol), 4-nitrobenzyl bromide (2.16 g, 10.0 mmol) and K₂CO₃ (1.38 g, 10.0 mmol) in CHCl₃ (30 mL) was stirred and refluxed for 4 h. After cooling the inorganic salt was filtered off, the organic phase was washed with water (10 mL), separated, dried (MgSO₄), filtered and evaporated. After flash column purification of the residue compound **22** was received as a yellow solid 2.53 g (68%), mp 107–108 °C. *R_f*: 0.48 (CHCl₃–MeOH, 9 : 1). Anal. calc. for C₂₀H₂₉N₄O₃: C 64.30, H 7.87, N 15.01; found: C 64.25, H 7.71, N 14.91%. MS (EI): *m/z* (%) 373 (M⁺, 65), 343(14), 234 (100), 221 (91), 136 (33).

General procedure for synthesis of compounds 12C–18C

To a stirred solution of amine **5–11** (3.0 mmol) and Et₃N (666 mg, 6.6 mmol) in CH₂Cl₂ (20 mL) was added dropwise a solution of freshly prepared 2-(chloroseleno)benzoyl chloride **4** (762 mg, 3.0 mmol) in CH₂Cl₂ (10 mL) over 5 min at rt and the mixture was stirred for a further 1 h. The organic phase was washed with brine (10 mL), dried (MgSO₄), filtered, evaporated and the residue was purified by flash column chromatography (CHCl₃–Et₂O or CHCl₃–MeOH) to give compounds **12C–18C** as orange-yellow solids in 35–73% yield.

2-[(1-Oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-carboxamidoprop)-3yl]benzo[d]isosenazol-3-one radical (12C). Yellow solid 541 mg (45%), mp 126–128 °C. *R_f*: 0.46 (CHCl₃–MeOH, 9 : 1). Anal. calc. for C₁₉H₂₄N₃O₃Se: C 54.16, H 5.74, N 9.97; found: C 54.02, H 5.68, N 9.82%. MS (EI): *m/z* (%) 424/422/420/419/418/416 (M⁺, 0.2/1/0.5/0.2/0.2/0.02), 394/392/390/389/388/386 (1/7/3/1/1/0.1), 186/184/182/181/180/179 (8/43/21/7/8/1), 136 (100). ⁷⁷Se NMR: 882 ppm.

2-[3-(1-Oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-yl)phenyl]benzo[d]isosenazol-3-one radical (14C). Yellow solid, 432 mg (35%), mp 148–150 °C. *R_f*: 0.57 (CHCl₃–Et₂O, 2 : 1). Anal. calc. for C₂₁H₂₁N₂O₂Se: C 61.17, H 5.13, N 6.79; found: C 61.10, H 5.12, N 6.67%. MS (EI): *m/z* (%) 415/413/411/410/409/407 (M⁺, 2/9/4/1/2/0.3), 385/383/381/380/379/377 (4/20/9/3/4/0.4), 199 (100), 184 (47).

2-[4-[1-(1-Oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-ylmethyl)piperazin-4-yl]phenylmethyl]benzo[d]isosenazol-3-one radical (15C). Yellow solid, 880 mg 56%, mp 115–117 °C. *R_f*: 0.34 (CHCl₃–MeOH, 9 : 1). Anal. calc. for C₂₇H₃₃N₄O₂Se: C 54.16, H 5.74, N 9.97; found: C 54.02, H 5.68, N 9.82%. MS (EI):

m/z (%) 527/525/523/522/521/519 (M⁺, 0.2/1/0.5/0.2/0.2/0.02), 497/495/493/492/491/489 (0.7/4/2/0.6/0.7/0.1), 290/288/286/285/284/282 (7/40/19/6/7/1), 136 (72), 122 (100).

2-(1-Oxyl-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-ylmethyl) benzo[d]isosenazol-3-one radical (16C). Pale yellow solid, 854 mg (61%), mp 165–166 °C. *R_f*: 0.32 (CHCl₃–Et₂O, 2 : 1). Anal. calc. for C₁₆H₁₉N₂O₂Se: C 54.86, H 5.47, N 8.00; found: C 54.71, H 5.44, N 7.93%. MS (EI): *m/z* (%) 353/351/349/348/347/345 (M⁺, 0.7/4/2/0.6/0.7/0.1), 323/321/319/318/317/315 (1.5/8/4/1/1.5/0.2), 138 (41), 122 (62), 107 (100).

2-(1-Oxyl-3-carbomethoxy-2,2,5,5-tetramethylpyrrolidine-4-ylmethyl) benzo[d]isosenazol-3-one radical (17C). Orange solid, 492 mg (40%), mp 207–209 °C. *R_f*: 0.26 (CHCl₃–MeOH, 9 : 1). Anal. calc. for C₁₈H₂₃N₂O₂Se: C 52.42, H 5.87, N 6.80; found: C 52.22, H 5.84, N 6.61%. MS (EI): *m/z* (%) 413/411/409/408/407/405 (M⁺, 1/5/3/0.8/1/0.1), 383/381/379/378/377/375 (0.7/4/2/0.6/0.7/0.1), 269/267/265/264/263/261 (7/40/19/6/7/1), 201/199/197/196/195/193 (18/100/47/15/18/2).

2-(1-Oxyl-2,2,6,6-tetramethylpiperidin-4-yl) benzo[d]isosenazol-3-one radical (18C). Orange-pink solid, 770 mg (73%), mp 230–231 °C. *R_f*: 0.44 (CHCl₃–Et₂O, 2 : 1). Anal. calc. for C₁₆H₂₁N₂O₂Se: C 54.38, H 5.99, N 7.93; found: C 54.30, H 5.88, N 7.85%. MS (EI): *m/z* (%) 355/353/351/350/349/347 (M⁺, 1/5/3/0.8/1/0.1), 186/184/182/181/180/178 (4/20/10/3/4/0.4), 140 (100).

General procedure for synthesis of compounds 12B, 13B, 15B, 18B

A solution of compound **12C**, **13C**, **15C** or **18C** (0.5 mmol) was refluxed with EtOH (15 mL) (saturated with HCl) for 20 min. After cooling, the solvent was evaporated off and the residue was crystallized with acetone or Et₂O to give title compounds as off-white or yellow solids in 68–82% yield.

2-[(1-Hydroxy-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-carboxamidoprop)-3yl]benzo[d]isosenazol-3-one hydrochloride (12B). Off-white solid 160 mg (70%), mp 115–117 °C. Anal. calc. for C₁₉H₂₆ClN₃O₃Se: C 49.74, H 5.71, N 9.16; found: C 49.72, H 5.70, N 8.99%. ¹H NMR (400 MHz, D₂O): δ 7.76 (m, 2H, ar CH), 7.56 (t, *J* = 6 Hz, 1H, ar CH), 7.39 (t, *J* = 6 Hz, 1H, ar CH), 6.05 (s, 1H, olefinic CH), 3.81 (t, *J* = 6.4 Hz, 2H, NCH₂), 3.26 (t, *J* = 6 Hz, 2H, NCH₂), 1.92 (m, 2H, CH₂–CH₂–CH₂), 1.45 (s, 6H, CCH₃), 1.32 (s, 6H, CCH₃). ¹³C NMR (100.5 MHz, D₂O): δ 168.7, 164.3, 139.5, 137.1, 135.1, 132.8, 127.8, 127.1, 126.8, 125.0, 78.6, 76.0, 43.41, 37.65, 28.6, 23.3 br, 22.07 br. ⁷⁷Se NMR (CDCl₃–MeOH): δ 1043; (D₂O): δ 925 ppm.

2-[3-(1-Hydroxy-2,5,5-trimethylpyrrolidin-2-yl)phenyl]benzo[d]isosenazol-3-one hydrochloride (13B). White solid 148 mg (68%), mp 141–142 °C. Anal. calc. for C₂₀H₂₃ClN₂O₂Se: C 54.87, H 5.29, N 6.40; found: C 54.73, H 5.20, N 6.22%. ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.14 (d, *J* = 7.7 Hz, 1H, ar CH), 7.88 (d, *J* = 8.0 Hz, 1H, ar CH), 7.87 (s, 1H, ar CH), 7.66 (dd, *J*₁ = 7.7 Hz, *J*₂ = 7.4 Hz, 2H, ar CH), 7.56 (d, *J* = 7.7 Hz, 1H, ar CH), 7.51 (d, *J* = 7.5 Hz, 1H, ar CH), 7.47 (dd, *J*₁ = 7.5 Hz, *J*₂ = 7.4 Hz, 2H, ar CH), 2.58 (m, 1H, CH₂), 2.37 (m, 1H, CH₂), 2.21 (m, 1H, CH₂), 2.04 (m, 1H, CH₂), 1.63 (s, 3H, CCH₃), 1.41 (s, 3H, CCH₃), 1.28 (br s, 3H, CCH₃). ¹³C NMR (100.5 MHz, D₂O): δ 168.1, 140.0, 138.3, 133.3, 130.4, 128.3, 127.4, 127.0, 126.6, 125.0, 76.6, 75.1, 35.1, 31.4 ppm. ⁷⁷Se NMR (76.3 MHz, DMSO-*d*₆): δ 891 ppm.

2-[4-[1-(1-Hydroxy-2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-ylmethyl)piperazin-4-yl]phenylmethyl]benzo[d]isosenazol-3-one trihydrochloride (15B). White solid, 286 mg (82%), mp 195–197 °C. Anal. calc. for C₂₇H₃₇Cl₃N₄O₂Se: C 51.08, H 5.87,

N 8.82; found: C 51.03, H 5.79, N 8.70%. ¹H NMR (400 MHz, D₂O): δ 7.96 (d, *J* = 7.6 Hz, 1H, ar CH), 7.89 (d, *J* = 7.2 Hz, 1H, ar CH), 7.72 (dd, *J*₁ = 7.6 Hz, *J*₂ = 7.2 Hz, 1H, ar CH), 7.59 (br s, 4H, ar CH), 7.53 (dd, *J*₁ = 7.6 Hz, *J*₂ = 7.2 Hz, 1H, ar CH), 5.84 (s, 1H, olefinic CH), 4.41 (s, 2H, NCH₂), 4.35 (s, 2H, NCH₂), 3.51 (s, 2H, NCH₂), 3.34 (brs, 2H, NCH₂), 1.47 (s, 6H, CCH₃), 1.45 (s, 6H, CCH₃). ¹³C NMR (100.5 MHz, D₂O): δ 167.9, 140.1, 133.3, 132.7, 127.0, 126.8, 125.0, 78.5, 76.8, 57.7, 49.6, 49.0, 24.2 br, 22.9 ppm. ⁷⁷Se NMR (76.3 MHz, D₂O): δ 997 ppm.

2-(1-Hydroxy-2,2,6,6-tetramethylpiperidin-4-yl) benzo[d]-isosenazol-3-one radical hydrochloride (18B). Yellow solid, 144 mg (74%), mp: 205–206 °C. Anal. calc. for C₁₆H₂₃ClN₂O₂Se: C 49.30, H 5.95, N 7.19; found: C 49.22, H 5.92, N 7.03%. ¹H NMR (400 MHz, D₂O): δ 7.87 (d, *J* = 7.6 Hz, 1H, ar CH), 7.82 (d, *J* = 8.0 Hz, 1H, ar CH), 7.62 (dd, *J*₁ = 8.0 Hz, *J*₂ = 7.2 Hz, 1H, ar CH), 7.46 (dd, *J*₁ = 7.6 Hz, *J*₂ = 7.2 Hz, 1H, ar CH), 4.93 (m, 1H, NCH), 2.30 (m, 2H, CHH), 2.10 (n, 2H, CHH), 1.52 (s, 6H, CCH₃), 1.43 (s, 6H, CCH₃). ¹³C NMR (100.5 MHz, D₂O): δ 168.7, 139.5, 132.9, 127.8, 127.4, 126.9, 125.0, 68.8, 45.0, 42.4, 27.3, 19.6 ppm. ⁷⁷Se NMR (76.3 MHz, D₂O): δ 890 ppm.

General procedure for synthesis of compounds 12A–18A

To a solution of nitroxide **12C–18C** (1.0 mmol) in AcOH (8 mL) Fe powder (560 mg, 10 mmol) was added and the mixture was warmed up to 70 °C until the reaction started. The mixture was stirred at room temperature for 1 h, diluted with water (15 mL), decanted and the decanted aqueous solution made alkaline with solid K₂CO₃. The mixture was extracted with CHCl₃ (3 × 15 mL), dried (MgSO₄), filtered, evaporated and after chromatographic purification (CHCl₃–MeOH) we got the title amines **12A–18A** in 47–59% yield.

2,2'-Diselenobis[N-(2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-carboxamido-prop-3-yl)]benzamide (12A). Beige solid, 199 mg (49%), mp 182–183 °C (2HCl salt). *R*_f: 0.16 (MeOH). Anal. calc. for C₃₈H₅₄Cl₂N₆O₄Se₂: C 51.42, H 6.13, N 9.47; found: C 51.30, H 6.00, N 9.31%. ¹H NMR (400 MHz, D₂O): δ 7.47 (d, *J* = 7.6 Hz, 2H), 7.48 (d, *J* = 7.6 Hz, 2H), 7.03 (dd, *J*₁ = 7.6 Hz, *J*₂ = 7.2 Hz, 2H), 6.97 (dd, *J*₁ = 7.6 Hz, *J*₂ = 7.2 Hz, 2H), 6.15 (s, 2H), 3.31 (br dd, 4H), 3.22 (br dd, 4H), 1.75 (br dd, 4H), 1.53 (s, 6H), 1.41 (s, 6H). ¹³C NMR (100.5 MHz, D₂O): δ 170.0, 164.6, 138.1, 136.6, 133.6, 132.2, 131.3, 127.9, 127.2, 71.8, 68.8, 37.8, 37.3, 28.3, 26.5, 26.2 ppm. ⁷⁷Se NMR (CDCl₃–MeOH): δ 442 ppm; (D₂O): δ 449 ppm.

2,2'-Diselenobis[N-3-(2,5,5-trimethylpyrrolidin-2-yl)phenyl]-benzamide oxalate (13A). White solid, 474 mg (55%), mp 161–164 °C. *R*_f: 0.11 (CHCl₃–MeOH, 2 : 1). Anal. calc. for C₄₂H₄₈N₄O₆Se₂: C 58.47, H 5.61, N 6.49; found: C 58.33, H 5.87, N 6.40%. ¹H NMR (for base) (400 MHz, CDCl₃): δ 9.9 (br s, 2H, NH), 7.92 (s, 2H, ar CH), 7.80 (d, *J* = 6.4 Hz, 2H, ar CH), 7.76 (d, *J* = 7.6 Hz, 2H, ar CH), 7.22–7.28 (m, 4H, ar CH), 7.11–7.18 (m, 6H, ar CH), 2.44–2.52 (m, 2H, CH₂), 2.08–2.18 (m, 2H, CH₂), 1.95–2.02 (m, 2H, CH₂), 1.82–1.90 (m, 2H, CH₂), 1.69 (s, 6H, CCH₃), 1.21 (s, 6H, CCH₃), 1.12 (s, 6H, CCH₃). ¹³C NMR (100.5 MHz, CDCl₃): δ 167.2, 143.5, 139.2, 133.9, 133.3, 132.0, 131.3, 129.4, 128.6, 126.3, 121.7, 120.8, 118.7, 70.05, 65.0, 38.3, 36.4, 29.20, 29.15, 28.6. ⁷⁷Se NMR (D₂O): δ 455 ppm.

2,2'-Diselenobis[N-3-(2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-yl)phenyl]benzamide (14A). Yellow solid, 398 mg (50%), mp 217–220 °C. *R*_f: 0.50 (CHCl₃–MeOH, 2 : 1). Anal. calc. for C₄₂H₄₆N₄O₂Se₂: C 63.31, H 5.85, N 7.03; found: C 63.27, H 5.76, N 7.00%. ¹H NMR (400 MHz, DMSO-d₆): δ 8.51 (d, *J* = 8.0 Hz, 1H), 7.82 (d, *J* = 6.8 Hz, 1H), 7.78 (s, 1H), 7.57 (m, 1H), 7.50 (m, 1H), 7.40 (m, 2H), 7.22 (d, *J* = 7.2 Hz, 1H), 6.07 (s, 1H), 2.90 (s, 1H), 1.65 (s, 6H), 1.54 (s, 6H). ¹³C NMR

(100.5 MHz, DMSO-d₆): δ 165.1, 142.9, 140.9, 139.9, 132.9, 131.5, 131.3, 129.9, 129.3, 127.3, 127.1, 125.7, 124.0, 123.8, 122.8, 70.5, 66.7, 27.1 ppm. ⁷⁷Se NMR (DMSO-d₆): δ 448 ppm.

2,2'-Diselenobis[N-{4-[1-(2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-ylmethyl)piperazin-4-yl]phenylmethyl}]benzamide (15A). Yellow solid, 479 mg (47%), mp 112–115 °C. *R*_f: 0.20 [MeOH–NH₄OH (aq. 25%), 40 : 1]. Anal. calc. for C₅₄H₇₀N₈O₂Se₂: C 63.52, H 6.91, N 10.97; found: C 63.35, H 7.02, N 10.88%. ¹H NMR (400 MHz, DMSO-d₆): δ 10.5 (br, 2H), 7.92 (d, *J* = 6.0 Hz, 2H), 7.78 (br, 2H), 7.67 (d, *J* = 6.8 Hz, 2H), 7.39 (m, 4H), 7.26 (m, 4H), 5.36 (s, 2H), 3.49 (br, 8H), 3.41 (s, 4H), 2.83 (s, 4H), 2.35 (br, 8H) 1.13 (s, 6H), 1.10 (s, 6H). ¹³C NMR (100.5 MHz, DMSO-d₆): δ 166.1, 142.9, 137.5 br, 133.8 br, 133.2, 131.9, 129.1, 128.5, 126.3, 120.3, 65.9, 62.6, 61.6, 54.9, 53.0, 52.6, 45.6, 30.9, 29.7, 21.7, 11.6 ppm. ⁷⁷Se NMR (DMSO-d₆): δ 448 ppm.

2,2'-Diselenobis[N-(2,2,5,5-tetramethyl-2,5-dihydro-1H-pyrrol-3-ylmethyl)]benzamide (16A). White solid, 349 mg (52%), mp 196–197 °C. *R*_f: 0.87 (MeOH–NH₄OH, 40 : 1). Anal. calc. for C₃₂H₄₂N₄O₂Se₂: C 57.14, H 6.29, N 8.33; found: C 57.11, H 6.15, N 8.20%. ¹H NMR (400 MHz, DMSO-d₆): δ 8.90 (br, 2H), 7.80 (d, *J* = 6.4 Hz, 2H), 7.68 (d, *J* = 6.8 Hz, 2H), 7.31 (m, 4H), 5.40 (s, 2H), 3.93 (br s, 4H), 1.19 (s, 12H), 1.11 (s, 12H). ¹³C NMR (100.5 MHz, DMSO-d₆): δ 167.0, 143.9, 133.1, 132.0, 131.4, 131.1, 129.8, 127.9, 126.1, 65.4, 62.6, 36.1, 31.1, 29.8 ppm. ⁷⁷Se NMR (DMSO-d₆): δ 447 ppm.

2,2'-Diselenobis[N-(3-carbomethoxy-2,2,5,5-tetramethylpyrrolidine-4-ylmethyl)]benzamide (17A). Beige solid, 443 mg (56%), mp 207–209 °C. *R*_f: 0.67 (MeOH–NH₄OH, 40 : 1). Anal. calc. for C₃₆H₅₀N₄O₄Se₂: C 54.54, H 6.36, N 7.07; found: C 54.50, H 6.20, N 6.95%. ¹H NMR (400 MHz, CDCl₃): δ 7.85 (d, *J* = 8.0 Hz, 2H), 7.49 (d, *J* = 7.2 Hz, 2H), 7.20 (m, 4H), 7.04 (br s, 2H), 3.64 (s, 6H, OCH₃), 3.56–3.59 (m, 2H), 3.36 (m, 2H), 2.80 (d, 2H), 2.51 (m, 6H) 1.38 (s, 6H), 1.28 (s, 6H), 1.09 (s, 6H), 1.04 (s, 6H). ¹³C NMR (100.5 MHz, CDCl₃): δ 175.0, 167.8, 133.5, 132.2, 131.6, 131.2, 126.5, 125.9, 60.1, 59.93, 59.86, 52.1, 50.5, 40.8, 31.7, 29.7, 27.4, 25.1 ppm. ⁷⁷Se NMR (CDCl₃): δ 459.6, 459.8 ppm (because of diastereomers).

2,2'-Diselenobis[N-(2,2,6,6-tetramethylpiperidin-4-yl)]benzamide (18A). White solid, 354 mg (49%), mp 224–226 °C. *R*_f: 0.35 (MeOH–NH₄OH, 40 : 1). Anal. calc. for C₃₆H₄₆N₄O₂Se₂: C 59.66, H 6.40, N 7.73; found: C 54.60, H 6.38, N 7.67%. ¹H NMR (400 MHz, DMSO-d₆): δ 8.42 (br, 2H), 8.0 (d, *J* = 6.4 Hz, 2H), 7.68 (d, *J* = 6.8 Hz, 2H), 7.31 (m, 4H), 5.40 (s, 2H), 3.93 (br s, 4H), 1.19 (s, 12H), 1.11 (s, 12H). ¹³C NMR (100.5 MHz, DMSO-d₆): δ 167.0, 143.9, 133.1, 132.0, 131.4, 131.1, 129.8, 127.9, 126.1, 65.4, 62.6, 36.1, 31.1, 29.8 ppm. ⁷⁷Se NMR (DMSO-d₆): δ 447 ppm.

Acknowledgements

This study was supported by the Hungarian Research Foundation (OTKA grant T048334 and M045190), the Deutsche Forschungsgemeinschaft (Si255/11-1) and partly by the Alexander von Humboldt-Stiftung in the form of a research fellowship to G. M. The authors thank to Dr J. Jek (ICN Hungary) for helpful discussion. H. S. is a Fellow of the National Foundation for Cancer Research (NFCR), Bethesda, MD, USA.

References

- 1 T. C. Stadtman, *J. Biol. Chem.*, 1991, **266**, 16257.
- 2 F. Ursini, in *Oxidative Processes and Antioxidants*, ed. R. Paoletti, New York, Raven Press, 1994, p. 25.
- 3 L. Flohé, E. A. Günzler and H. H. Schock, *FEBS Lett.*, 1973, **32**, 132.
- 4 J. T. Rotruck, A. L. Pope, H. E. Ganther, A. B. Swanson, D. G. Hafeman and W. G. Hoekstra, *Science*, 1973, **179**, 588.

- 5 M. Maiorino, K. D. Aumann, R. Brigelius-Flohé, D. Doria, J. van den Heuvel, J. McCarthy, A. Roveri, F. Ursini and L. Flohé, *Biol. Chem. Hoppe-Seyler*, 1995, **376**, 651.
- 6 C. Rocher, J. L. Lalanne and J. Chaudière, *Eur. J. Biochem.*, 1992, **205**, 955.
- 7 K. R. Maddipati and L. J. Marnett, *J. Biol. Chem.*, 1987, **262**, 17398.
- 8 F. F. Chu, J. H. Doroshov and R. S. Esworthy, *J. Biol. Chem.*, 1993, **268**, 2571.
- 9 R. Brigelius-Flohé, *Free Radical Biol. Med.*, 1999, **27**, 951.
- 10 C. Krebs-Remy, P. Mehlen, M. E. Mirault and A. P. Arrigo, *J. Cell Biol.*, 1996, **133**, 1083.
- 11 F. Weitzel and A. Wendel, *J. Biol. Chem.*, 1993, **268**, 6288.
- 12 G. Mugesh and H. B. Singh, *Chem. Soc. Rev.*, 2000, **29**, 347.
- 13 C. A. Collins, F. H. Fry, A. L. Holme, A. Yiakouvaki, A. Qenaï, C. Pourzand and C. Jacob, *Org. Biomol. Chem.*, 2005, **3**, 1541.
- 14 G. Mugesh, W. W. du Mont and H. Sies, *Chem. Rev.*, 2001, **101**, 2125.
- 15 A. Müller, E. Cadenas, P. Graf and H. Sies, *Biochem. Pharmacol.*, 1984, **33**, 3235.
- 16 A. Wendel, M. Fausel, H. Safayhi, G. Tiegs and R. Otter, *Biochem. Pharmacol.*, 1984, **33**, 3241.
- 17 H. J. Reich and C. P. Japsense, *J. Am. Chem. Soc.*, 1987, **109**, 5549.
- 18 P. Chan, P. Cotelle, L. J. Bernier and J. P. Henichart, *Bioorg. Med. Chem. Lett.*, 1991, **1**, 277.
- 19 P. V. Jacquemin, L. E. Christiaens, M. J. Renson, M. J. Evers and N. Dereu, *Tetrahedron Lett.*, 1992, **33**, 3863.
- 20 J. Chaudière, J. C. Yadan, I. Erdelmeier, C. Tailhan-Lomont, and M. Moutet, in *Oxidative Processes and Antioxidants*, ed. R. Paoletti, New York, Raven Press, 1994, p. 165.
- 21 A. Mohsine and L. Christiaens, L., *Heterocycles*, 1996, **43**, 2567.
- 22 T. G. Back and B. P. Dyck, *J. Am. Chem. Soc.*, 1997, **119**, 2079.
- 23 V. Galet, J. Bernier, J. Henichart, D. Lesieur, C. Abadie, L. Rochette, A. Lindenbaum, J. Chalas, J. Renaud, de la Faverie, B. Pfeiffer and P. Renard, *J. Med. Chem.*, 1994, **37**, 2903.
- 24 S. Ostrovidov, P. Franck, D. Joseph, L. Martarello, G. Kirsch, F. Belleville, P. Nabet and B. Dousset, *J. Med. Chem.*, 200, **43**, 1762.
- 25 I. A. Cotgreave, P. Moldeus, R. Brattsand, A. Hallberg, C. M. Andersson and L. Engman, *Biochem. Pharmacol.*, 1992, **43**, 793.
- 26 S. R. Wilson, P. A. Zucker, R. R. C. Huang and A. Spector, *J. Am. Chem. Soc.*, 1989, **111**, 5936.
- 27 M. Iwaoka and S. Tomoda, *J. Am. Chem. Soc.*, 1994, **116**, 2557.
- 28 T. Wirth, *Molecules*, 1998, **3**, 164.
- 29 G. Mugesh, A. Panda, H. B. Singh, N. S. Punekar and R. J. Butcher, *Chem. Commun.*, 1998, 2227.
- 30 G. Mugesh, A. Panda, H. B. Singh, N. S. Punekar and R. J. Butcher, *J. Am. Chem. Soc.*, 2001, **123**, 839.
- 31 H. Sies, *Free Radical Biol. Med.*, 1993, **14**, 313.
- 32 T. P. M. Akerboom, H. Sies and D. M. Ziegler, *Arch. Biochem. Biophys.*, 1995, **316**, 220.
- 33 J. Chaudière, O. Courtin and J. LeClaire, *Arch. Biochem. Biophys.*, 1992, **296**, 328.
- 34 H. O. Hankovszky, K. Hideg, I. Bódi and L. Frank, L., *J. Med. Chem.*, 1986, **29**, 1138.
- 35 M. C. Krishna, W. Degraff, H. O. Hankovszky, P. C. Sár, T. Kálai, J. Jek, A. Russo, J. B. Mitchell and K. Hideg, *J. Med. Chem.*, 1998, **41**, 3477.
- 36 R. A. Shankar, K. Hideg, J. L. Zweier and P. Kuppusamy, P., *J. Pharmacol. Exp. Ther.*, 2000, **292**, 838–845.
- 37 P. Twomey, J. Taira, W. DeGraff, J. B. Mitchell, A. Russo, M. C. Krishna, H. O. Hankovszky, L. Frank and K. Hideg, *Free Radical Biol. Med.*, 1997, **22**, 909.
- 38 S. Gadányi, T. Kálai, J. Jek, Z. Berente and K. Hideg, *Synthesis*, 2000, 2039.
- 39 R. Cantineau, G. Thiange, A. Pelenevaux, L. Christiaens and M. Guillaume, *J. Labelled Compd. Radiopharm.*, 1986, **23**, 59.
- 40 H. O. Hankovszky, K. Hideg, M. J. Lovas, G. Jerkovich, A. Rockenbauer, M. Gyr and P. Sohár, *Can. J. Chem.*, 1989, **67**, 1392.
- 41 H. O. Hankovszky, K. Hideg, K. and L. Lex, *Synthesis*, 1981, 147.
- 42 K. Hideg, H. O. Hankovszky, H. A. Halász and P. Sohár, *J. Chem. Soc., Perkin Trans. 1.*, 1988, 2905.
- 43 E. G. Rozantsev, *Free Nitroxyl Radicals*, New York, Plenum Press, 1970.
- 44 T. Kálai, M. Balog, J. Jek, W. L. Hubbell and K. Hideg, *Synthesis*, 2002, 2365.
- 45 S. Ram and R. E. Ehrenkauffer, *Synthesis*, 1988, 91.
- 46 T. Kálai, H. O. Hankovszky, É. Hideg, J. Jek and K. Hideg, *ARKIVOC*, 2002, **3**(iii), 112.
- 47 P. C. Sár, T. Kálai, M. N. Bárász, G. Jerkovich and K. Hideg, *Synth. Commun.*, 1995, **25**, 2929.
- 48 H. Fischer and N. Dereu, *Bull. Soc. Chim. Belg.*, 1987, **96**, 757.
- 49 A. Roveri, M. Maiorino and F. Ursini, *Methods Enzymol.*, 1994, **233**, 202.